# **Tropical Journal of Natural Product Research**

Available online at https://www.tjnpr.org

**Original Research Article** 



## Effect of *Cnidoscolus aconitifolius* Extract on Rat Liver Injury Induced with Carbon Tetrachloride (CCl<sub>4</sub>)

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## ARTICLE INFO

## ABSTRACT

Article history: Received 26 October 2022 Revised 10 November 2022 Accepted 11 November 2022 Published online 01 December 2022

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Cnidoscolus aconitifolius in Indonesia is consumed as a vegetable which contains phenolic compounds known to have activity in the liver. This study was designed to determine the hepatoprotective effect of various doses of C. aconitifolius extract on the liver of rats injury induced with carbon tetrachloride (CCl<sub>4</sub>). Thirty-six Wistar rats were used in this study, divided into six groups. Normal group, positive control, negative control, and treatment groups of C. aconitifolius extract (50, 100, and 200 mg/kg). The study analysed serum transaminases and liver histopathology of rats. Serum analysis was performed to determine the activity of aspartate aminotransferase (AST), alanine aminotransferase (ALT), total cholesterol, and albumin levels. The histopathology of rat liver was analyzed using the Mandja Roenigk method. This study showed that in the groups treated with C. aconitifolius extract at the doses of 50 mg/kg, 100 mg/kg and 200 mg/kg (Group D, E, F) respectively, there was a decrease in ALT, AST and cholesterol levels. In contrast, albumin levels increased compared to the positive control (Group B) (p<0.05). The administration of C. aconitifolius extract (Group F: 200 mg/kg b.w) had almost the same effect as the administration of Curcuma Force® (Group C) (P>0.05). This study showed improvement in the liver function of rats after administration of C. aconitifolius extract. In conclusion, C. aconitifolius possesses a hepatoprotective effect on rat liver damage caused by CCl₄ induction.

Keywords: Cnidoscolus aconitifolius, Hepatoprotective, Liver, Carbon tetrachloride.

## Introduction

Liver damage is common in patients who use synthetic drugs continuously.1 This can happen because the liver metabolizes most drugs which can reduce liver function. The use of natural materials is an alternative choice for solving these problems. Various types of plants have long been used in traditional medicine. Currently, many studies use plant samples to find the activity of active molecules that can be used as drug candidates in the future.<sup>2</sup> Cnidoscolus aconitifolius belongs to the group of arborescent shrubs. This plant can grow up to 6 meters, the leaves are lobed palmately green, and have small flowers in dichotomous branched cymes and milky sap. C.aconitifolius is often used as food, medicine, and ornamental plant until now.3 C. aconitifolius has several pharmacological effects such as hypoglycemic4, antibacterial3, antioxidant5, analgesic and antiinflammatory<sup>6</sup>, treatment of high-dose paracetamol poisoning<sup>7</sup>, and hepatoprotective.8 This plant was first cultivated as a vegetable in the regions of Southeastern Mexico and Maya Guatemala.9 Due to the easy cultivation process, the plant has spread throughout the world including the tropics. Indonesia as a tropical country has many types of plants that can be developed into treatment options. C. aconitifolius is a type of plant that grows well in Indonesia and it is still a prospect to observe its effects on human health further. Indonesian people often call it Japanese Papaya and are usually only used as a vegetable food ingredient.

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**Citation:** Muslim Z, Farizal J, Abdillah R, Sunita S. Effect of *Cnidoscolus aconitifolius* Extract on Rat Liver Injury Induced with Carbon Tetrachloride (CCl<sub>4</sub>). Trop J Nat Prod Res. 2022; 6(11):1809-1813. http://www.doi.org/10.26538/tjnpr/v6i11.11

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C. aconitifolius in Nigeria is known as Chaya and is usually consumed in Yoruba-land, Nigeria as a vegetable.<sup>10</sup>

*C. aconitifolius* leaves contain Tannins, Saponins, Alkaloids, Flavonoids, Cyanogenic Glycosides, and Phytates.<sup>11</sup> These benefits mainly come from the phenolic compounds it contains. The results of research on the therapeutic effect of *C. aconitifolius* as a hepatoprotector are still very limited, especially regarding variations in dose and liver function parameters of rats induced by carbon tetrachloride (CCl<sub>4</sub>). The liver enzyme alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (APT), albumin (Alb), total bilirubin, and total protein (TP) are biomarkers that can be observed to estimate the extent of hepatocyte damage.<sup>12</sup> This study aim to determine the hepatoprotective effect of *c. aconitifolius* on CCl<sub>4</sub>-induced rat liver function.

## **Materials and Methods**

## Plant Materials

*C. aconitifolius* was collected from Bengkulu Province, Indonesia. *C. Aconitifolius* was identified in the Biology Herbarium, Faculty of Mathematics and Natural Sciences, Bengkulu University, with certificate number 525/UN30.12.LAB.BIOLOGI/KM/2021.

#### Exstraction C. aconitifolius

*C. aconitifolius* leaves began with the selection of samples with the criteria of plants being more than one year old and choosing leaves that are not too old. Leaf samples collected were approximately 5 kg, dried to a moisture content of 6.41% and mashed. The extraction process was carried out at the Biota Sumatra Laboratory, Andalas University. A dried sample (500 g) was extracted by maceration method using 5 litre 70% ethanol. After the maceration process was completed, the filtrate was evaporated using a Rotary Evaporator (Buchi® R210), and 8 g of thick extract was obtained.

The extraction process was carried out according to the standards of the Indonesian Herbal Pharmacopoeia Edition 2.  $^{13}$ 

## Animals and experimental design

This study used male Wistar rats. The weight of the animals was 200-250 g. The study protocols were approved by the Health Research Ethics Commission of the Bengkulu Ministry of Health Poltekkes; the approval number KEPK/114/04/2022. The animal study was carried out at the Animal House, Faculty of Pharmacy at Andalas University. Rats were placed in separate cages for two weeks for acclimatization under laboratory conditions (humidity 50%  $\pm$  10%, temperature 27°C  $\pm$  3°C and under 12 h light - 12 h dark periods). Animals receive adequate food and drink according to standard procedures based on the Guide for the Care and Use of Laboratory Animals. All activities are carried out by minimizing animal pain and using animals as efficiently as possible.

#### Experimental Design

Experimental animals were divided into six groups that received intervention for two weeks. Each group consisted of 6 rats. Table 1 describes the groups of treatment in detail. All groups in this study were induced with CCL<sub>4</sub> (1.25 mL/kg) as a hepatotoxic agent obtained from the Laboratory of the Faculty of Pharmacy, Andalas University, except group A as normal control. Curcuma Force<sup>®</sup> (4 mg/kg) contains *Curcumae xanthorrhizae* and *Piperis nigris* produced by SOHO Pharmaceutical Industry as a positive control. The variuos doses of the *C. aconitifolius* extract used were 50 mg/kg; 100 mg/kg; and 200 mg/kg. CCl<sub>4</sub> (1.25 mL/kg) was administered via the intraperitoneal route while the natrium carboximetilcelulosa (CMC Na) suspension and extract were administered orally.

## Blood Sample Collection and Analysis

After the intervention for two weeks, blood was collected through the rat eye arteries as much as 5 mL using an EDTA tube. Rat blood analysis was performed using a Hematology Analyzer with DiaSys<sup>®</sup> reagents. Then the animal was sacrificed by cervical dislocation. Before the animals were sacrificed, Ketamine (40 mg/kg b.w) and Xylazine (5 mg/kg b.w) were administered intraperitoneally as anaesthetics and analgesics.

#### Histological Analysis

Livers of the rats were collected after the animals were sacrificed by cervical dislocation. Before sectioning, the liver was fixed in a 10% buffered formalin solution for 24 hours. The liver tissue was dehydrated in 70%, 80%, 90%, and 96% alcohol, xylol, and liquid paraffin, in that order. The following stages are tissue vacuuming and embedding. The liver tissue was sectioned and stained with hematoxylin and eosin at a thickness of 4-6 mm. Histological findings of the liver included central veins, hepatocytes, hepatocyte cell arrangement, and pathological liver abnormalities such as parenchymal degeneration, hydropic degeneration, and necrosis of liver hepatocytes. A 40-times magnification microscope with Olympus CX33<sup>®</sup> was used to examine liver tissue in five distinct fields of view. The average stained area and staining intensity of the five fields were chosen.

#### Table 1: Experimental groups

Groups	Treatment		
А	CMC Na 1%		
В	CMC Na 1% + CCl4*		
С	CMC Na 1% + CCl4* + Curcuma Force®*		
D	CCl4* + CMC Na 1% + 50mg/kg b.w extract of C. aconitifolius		
Е	CCl4* + CMC Na 1% + 100mg/kg b.w extract of C. aconitifolius		
F	CCl4* + CMC Na 1% + 200mg/kg b.w extract of C. aconitifolius		
* CCL4 d	loses: 1.25 mL/kg b.w		

\* CCL4 doses: 1.25 mL/kg b.w

\* Curcuma Force® doses: 4mg/kg b.w

A semi-quantitative histopathology score modified from the recently recognised Manja-Roegnik criteria was used to assess the histopathological characteristics of liver damage.

#### Statistical analysis

AST, ALT, Albumin and Total Cholesterol examination data were analyzed by the one-way ANOVA method using SPSS<sup>®</sup> software. Statistical analysis obtained the mean, standard deviation (SD) and P-Value. P-values at p<0.05 were considered significant.

## **Results and Discussion**

#### Effect of C. aconitifolius on rat liver function parameters

This study evaluated the effect of *C. aconitifolius* extract on CCl<sub>4</sub>induced rat liver damage. Liver cell damage can be caused by various factors such as autoimmune disease, bacteria, viruses, and parasites, the influence of chemicals, such as some drugs (high doses of paracetamol and antituberculosis drugs), toxic compounds (thioacetamide and carbon tetrachloride (CCl<sub>4</sub>)).<sup>14</sup> Research has led to the use of plants that have hepatoprotective effects. However, no substance is fully effective in providing complete protection to the liver, or helping to regenerate liver cells.<sup>15</sup>

The results of serum examination in group B showed damage to the liver cells of rats. These results were indicated by a decrease in serum albumin levels compared to other treatment groups, while total cholesterol levels and AST and ALT activities increased (Table 2). In group C administered Curcuma force<sup>®</sup> to rats and liver damage induced with CCl<sub>4</sub>, the results show a decrease in AST, ALT, and total cholesterol. At the same time, albumin levels were slightly increased compared to group B (p<0.05) (Table 2).

The findings in this study showed that there was liver damage caused by CCl<sub>4</sub> based on serum assays and histopathological analysis. Previous studies that used CCl<sub>4</sub> as an inducer of liver damage in experimental studies using animals, proved successful in damaging liver cells at a dose of 1.25 mL/kg b.w. CCl4 damages liver cells by activating enzymes such as CYP2E1, CYP2B1 or CYP2B2 and possibly CYP3A, which reduce CCl<sub>4</sub> to CCl<sub>3</sub>, which is a free radical.<sup>16</sup> CCl<sub>3</sub> causes oxidative stress in liver cells, thereby increasing the activity of transaminase enzymes and the formation of steatosis.<sup>17</sup> CCl<sub>4</sub> was also reported to reduce albumin levels in rats induced for ten weeks.<sup>18</sup> CCl<sub>4</sub> induction can increase the synthesis of fatty acids, triglycerides and cholesterol.<sup>19</sup> Based on the ability of CCl<sub>4</sub> to damage liver cells, CCl<sub>4</sub> was used as a negative control in this study.

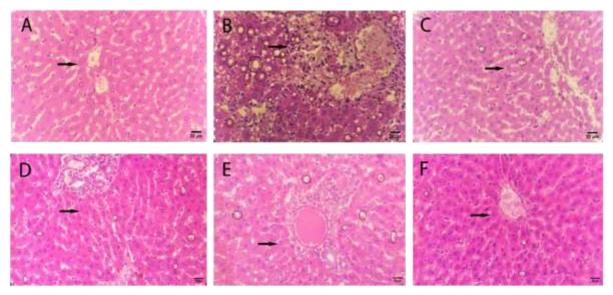
Curcuma is one of the natural ingredients that is well known as a hepatoprotective.<sup>20</sup> Previous studies reported that piperine is a potent inhibitor of CYP450 enzymes, particularly CYP3A4, CYP2C9, and CYP1A2.<sup>21</sup> CYPs also function in the biosynthesis of steroids, lipids, and other secondary metabolites.<sup>22</sup> In this study, we used Curcuma Force® containing *Curcumae xanthorrhizae* and *Piperis nigris* as positive controls. Decreased activity of the enzyme transaminase indicates that liver cells are undergoing repair. The increase in albumin levels to reach the normal range of albumin levels in rats also occurred in group C, which was treated with Curcuma Force<sup>®</sup>. Increased albumin levels are one sign of improvement in rat liver cells. The liver is an organ that functions for albumin synthesis.<sup>23</sup> Other studies suggest that Curcuma has the effect of lowering serum levels of lipid peroxides and total cholesterol.<sup>24</sup>

The results in groups D, E, and F showed that the activity of AST, ALT, and total cholesterol levels decreased. At the same time, albumin increased after administration of extract of *C. aconitifolius* (Table 2). The decreased activity was significant in AST and ALT (p<0,000), total cholesterol (p<0.001) and an increase in albumin (p<0.002). These results are linear with variations in the dose of *C. aconitifolius* extract. The higher the dose of *C. aconitifolius*, the better the hepatoprotective effect. Some research results state that *C. aconitifolius* contains phenolic compounds such as Hesperidin,<sup>25</sup> Kaempferol,<sup>26</sup> Lignin,<sup>25,26</sup> Protocatechic acid,<sup>25,29</sup> Quercetin,<sup>30</sup> Rutine.<sup>27,28</sup> Triterpenes, glycosides, phenolic compounds, and flavonoids as classes of compounds with hepatoprotective activity.<sup>31</sup> This study showed that *C. aconitifolius* ethanol extract had a good effect on the repair of rat liver cells damaged by CCl<sub>4</sub>.

Groups	AST (U/L)*	ALT (U/L)*	Albumin (g/dL)**	Total Cholesterol (mg/dL)***
А	$15.58 \pm 1,\!63$	$19.26 \pm 1.32$	$5.6\pm0.10$	$59.78 \pm 5.13$
В	$27.63 \pm 6.87^{\circ}$	$32.48 \pm 2.10^{c}$	$5.12\pm0.16^{\mathrm{c}}$	$73.61 \pm 6.59^{\circ}$
С	$19.95 \pm 3.18^{d}$	$24.3\pm1.34^{\ d}$	$5.17\pm0.28$	$59.66\pm5.89^{d}$
D	$25.75\pm1.53^{\text{b}}$	$31.35\pm0.98$	$5.11\pm0.33^{b}$	$65.83\pm6.80^{b}$
Е	$23.61\pm2.2^{b}$	$25.8 \pm 1.59^{ab}$	$5.39\pm0.38^{b}$	$65.2\pm2.76^{\rm b}$
F	$19.8\pm0.99^{\ ab}$	$24.6 \pm 1.31^{ab}$	$5.48\pm0.31^{b}$	$61.5\pm4.34^{ab}$

Table 2: AST, ALT, albumin and total cholesterol value post-treatment by extract of C. aconitifolius.

Values are presented as Mean  $\pm$  SD. There are 6 animals in each group (n=6). P < 0.005 value are considered statically significant. Asterisk \*P < 0.000; \*\*P < 0.002; \*\*\*P < 0.001. <sup>a</sup> indicates value is significantly different at P < 0.05 vs grup B. <sup>b</sup> indicates value is not significantly different at P > 0.05 vs grup C. <sup>c</sup> indicates value is not significantly different at P > 0.05 vs grup A. <sup>d</sup> indicates value is significantly different at P > 0.05 vs grup B. Statistical test between groups using one-way ANOVA and a Tukey's post hoc multiple comparison test.



**Figure 1:** Representative histopathological rat liver with magnification 400X. A (Group A: Normal Control) showed normal liver cells (arrows); B (Group B: Negative Control) showed the occurrence of necrosis (arrows) and steatosis (circles); C (Group C: Positive Control) showed that there were cell repairs in the form of parenchymal degeneration (arrows) to normal and steatosis (circles); D (Group D: Treatment 50 mg/kg b.w extract of *C. aconitifolius*) and E (Group E: 100 mg/kg b.w extract of *C. aconitifolius*) showed necrosis to hydrophilic degeneration (arrows) and (circles); F (Group F: 200 mg/kg b.w extract of *C. aconitifolius*) showed cell recovery (arrows) and reduced steatosis.

Table 3: Analysis Mandja Roenigk's histopatology score

Group*	Score**	p-Value***
А	$1\pm0.00$	
В	$4\pm0.00$	
С	$3\pm0.75$	0.011
D	$3.77\pm0.33$	
Е	$3.56 \pm 0.24$	
F	$3.3\pm0.73$	

\*Each group 5 repetation

\*\*Score are Mean  $\pm$  SD.

The higher the dose of *C. aconitifolius*, the more significant the improvement in liver cell condition based on serum and histopathological examinations.

The results of enzyme activity; AST, ALT, and total cholesterol levels appeared to decrease. In contrast, albumin levels increased in groups D, E, and F. This condition explained a significant difference compared to group B as a negative control (p<0.05), but there was no significant difference. A significant difference with group C as a positive control (p>0.05). Damage to hepatocytes, causing the release

of ALT and AST in the cytoplasm or mitochondria into the blood circulation. The ALT enzyme is located mainly in the hepatocyte cytosol. Meanwhile, AST enzymes are found not only in liver cells but in cells in other organs, such as the heart, kidneys, and pancreas. Increased ALT and AST activity is a sign of liver cell damage.<sup>32</sup> In conditions of acute liver injury, ALT increased more significantly than AST.<sup>33</sup>

#### Effect of C. aconitifolius on histophatologi of rat liver

Based on the histological observations of group B (Figure 1.B), necrosis of hepatocytes (arrows in Figure 1.B) and steatosis (circles in Figure 1.B) were seen. The results of Mandja Roenigk scoring of the histological samples in group B showed an average score of 4 (Table 3). The interpretation of Score 4 is that many cells undergo necrosis. Meanwhile, histopathological observations in group C showed improvement in liver cells to normal again (arrows in Figure 1.C), and steatosis appeared to be significantly reduced. The histopathological observation of rat liver (Figure 1. D, E, F) also showed improvement of liver cells with reduced necrosis starting from low doses (Group D) to normal hepatocytes seen at the highest dose of an extract of *C. aconitifolius*, the better the liver condition of the rats in this study. Mandja Roenigk scoring in groups C, D, E, and F showed improvement with a score of 3 (hydropic degeneration) (Table 3).

Based on the scoring given to each group, there was a significant difference between group B as a negative control and the other groups (p<0.05). Hepatocyte damage can be in the form of necrosis, parenchymal necrosis, inflammation, atypia, degeneration, hepatocyte pleomorphism, ductal proliferation, cytoplasmic eosinophilia, and inflammatory cells in the portal area.<sup>34</sup> Histopathological liver damage can be observed through microscopic examination in the form of cell swelling (hydropic degeneration) and fatness (steatosis).<sup>23</sup> In this study, we looked at the condition of rat liver cells through histopathological examination induced by CCl<sub>4</sub>, Curcuma Force<sup>®</sup>, and treated with *C. aconitifolius*.

Histopathological observations in this study used the Mandja Roenigk scoring. Normal cells were given a score of 1 point, cells with parenchymatous degeneration were given a score of 2 points, cells with hydropic degeneration were given a score of 3 points, and necrotic cells were given a score of 4 points.<sup>35</sup> Based on histopathological observations, Group A showed normal liver cell shape because it was only treated with 1% Na CMC (Figure1.A). The average score of Group A is 1 and can be interpreted as a normal hepatocyte condition. Microscopic observation of the liver specimens of Group B rats showed necrosis of hepatocytes and steatosis (Figure 1.B). Steatosis occurs due to the effect of CCl<sub>4</sub>, which has been reduced by Cytochrome P450 (CYP) enzymes.<sup>36</sup> Based on the results of microscopic observations, the average score of Group B was 4 (Table 4). So it can be interpreted that most hepatocytes in Group B are necrotic. The histopathological condition of steatosis decreased in Groups D, E and F, which was confirmed by a downward trend in the average total cholesterol level compared to the other groups (Table 2). The average score of hepatocyte condition in the group given the extract of C. aconitifolius was at a score of 3 (hydropic degeneration). Based on this, it can be seen that there is an improvement in the condition of the liver cells of rats given C. aconitifolius compared to the normal and negative control groups, while when compared to the group given Curcuma Force®, the improvement is the same with an average score of 3.

## Conclusion

Based on the results of serum analysis and confirmation of histopathological examination of the liver of rats showed that *C. aconitifolius* had a hepatoprotective effect. At a dose of 200mg/kg b.w, the hepatoprotective effect of the extract was comparable to the hepatoprotective effect of the positive control.

## **Conflict of Interest**

The authors declare no conflict of interest.

## **Authors' Declaration**

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

## Acknowledgements

The authors thank the Ministry of Health Republic of Indonesia for financial support. Faculty of Pharmacy, Universitas Andalas, and Balai Veteriner Bukit Tinggi, Indonesia for their technical assistance and support during this research.

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